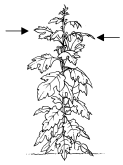
Crop Guide KB Item 3451v4

Chrysanthemum (G/H)

Sampling Notes

The nutritional status of this flower is monitored using soil tests and plant analysis. Monitoring regularly is important to help sustain optimum levels and avoid nutritional disorders. If disorders do occur, rapid diagnosis is necessary to assist correction.

Leaf



Sampling Time: From bud start to harvest.

Plant Part 4th leaf from the tip. Omit unfurled leaves.

Collect From: -

Quantity per Sample: 20-30 leaves.

Recommended Tests: Basic Plant (BP).

Comments: Just prior to flowering or during early flowering is considered to be the appropriate time to sample. However, regular leaf analyses should be obtained through the growing season (e.g. 6 - 8 weekly intervals) in order to monitor the effect of liquid feeding programmes being used.

Soil

Sampling Time: Prior to crop establishment.

Core Depth 15cm.

Collect From: From the rooting zone of the plants.

Quantity per Sample: 12 - 20 cores.

Recommended Tests: Basic Soil (BS), Soluble Salts (SSg).

Comments: If a problem is suspected during the growing season, then a sample should be taken from the rooting zone immediately adjacent to the plant. Collecting a second sample from an unaffected area may help identify the cause of the problem.

CHRYSANTHEMUM (G/H)

Comments

Ammonia toxicity has been observed in chrysanthemums where ammonium based fertilisers were used on high pH (>7) soils. The free ammonia damages the root hairs, reduces growth rates in the plant, and in severe cases eventually causes death of the plant.

The potassium content declines as the plant matures, especially during flower formation.

Magnesium deficiencies are less common in chrysanthemums than in other major glasshouse crops.

Both manganese deficiency and toxicity can arise in chrysanthemums.

Results for copper, zinc and manganese in leaves sprayed with fungicides will not be reliable due to adhering spray residues on the leaves.

Iron deficiencies have been observed, but symptoms may exist even when leaf levels appear satisfactory. This may be due to the presence of physiologically inactive forms of iron within the tissue. Also, soil contamination of leaves growing near the ground may elevate total iron results.

References

Bunt, A.C. 1976. Modern potting compost. George Allen and Unwin, p 129.

Scaife, A. and Turner, M. 1983. Diagnosis of mineral disorders in plants. Volume 2, Vegetables. MAFF/ARC London.

Cresswell, G.C. and Weir, R.G. 1997. Plant nutrient disorders 5. Ornamental plants and shrubs. Inkata Press.

Disclaimer

Normal Range levels shown as histograms in test reports relate specifically to the sampling procedure provided in this crop guide. The Normal Range levels in test reports and Comments provided in this Crop Guide are the most up to date available, but may be altered without notification. Such alterations are implemented immediately in the laboratory histogram reports. It is recommended that a consultant or crop specialist be involved with interpretations and recommendations.